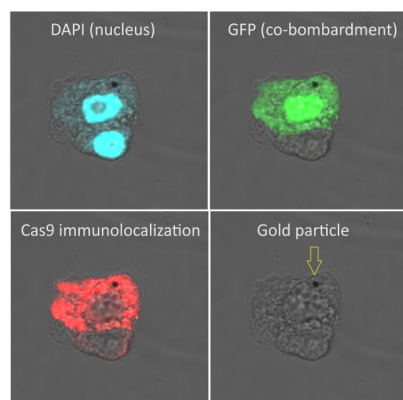


Product no AS11 1782**Goat anti-Rabbit IgG (H&L), DyLight® 550 conjugated, min, cross-reactivity to bovine,goat,human, mouse,Rat IgG****Product information**

Host	Goat
Clonality	Polyclonal
Purity	Immunogen affinity purified IgG.
Format	Lyophilized
Quantity	1 mg
Reconstitution	For reconstitution add 1,1 ml of sterile water, Let it stand 30 minutes at room temperature to dissolve, Prepare fresh working dilutions daily
Storage	Store lyophilized material at 2-8°C. Product is stable for 4 weeks at 2-8°C after rehydration. For long time storage after reconstitution, dilute the antibody solution with glycerol to a final concentration of 50% glycerol and store as liquid at -20°C, to prevent loss of enzymatic activity. For example, if you have reconstituted 1 mg of antibody in 1,1 ml of sterile water add 1,1 ml of glycerol. Such solution will not freeze in -20°C, If you are using a 1:5000 dilution prior to diluting with glycerol, then you would need to use a 1:2500 dilution after adding glycerol. Prepare working dilution prior to use and then discard. Be sure to mix well but without foaming.
Additional information	This antibody reacts with the heavy (gamma) chains on rabbit IgG and with the light chains on all rabbit immunoglobulins based on immunoelectrophoresis. Minimum cross-reactivity is observed to non-immunoglobulin rabbit serum proteins and bovine,goat,human,mouse, rat IgG based on immunoelectrophoresis. Antibody is supplied in 10 mM sodium phosphate, 150 mM sodium chloride, pH 7.2, 1 % (w/v) BSA, Protease/IgG free and 0.05 % (w/v) sodium azide as preservative.

Application information**Recommended dilution** 1 : 50 -1 : 5000 (ICC), 1 : 20 -1 : 2000 (IHC),**Application example**

Immunolocalization of Cas9 on maize (*Zea mays*, cv. H1233) plant suspension cultures. Rabbit anti-Cas9 antibody (Agrisera AS16 3690), diluted 1:100 and DyLight 550 anti-rabbit secondary antibody (Agrisera AS11 1782) diluted 1:300, (red) are used for immunodetection following formaldehyde fixation, cell wall digestion and detergent permeabilization of cells. DAPI is used as nuclear marker (blue). Gold particle used for microprojectile bombardment of plasmids (GFP and Cas9) is indicated with an arrow at the last column. Scalebar is 5µm.

Courtesy of Dr. Ferhan Ayaydin, Hungarian Academy of Science, Hungary

Labeling and detection protocol

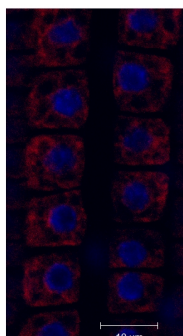
Fixation (30 min) 4% paraformaldehyde in PBS (pH 7.4) with 0.01% TritonX-100. (3x5min PBS wash)
 Cell wall digestion (40 min): 1% Cellulase, 1% Pectinase in 0.5% (w/v) MES buffer (pH 5.6) (2x5 min PBS wash)
 Immobilization of cells (10mins): Cells in PBS were settled onto poly-L-Lysine coated coverslips, excess PBS removed without air drying the cells.

This product is **for research use only** (not for diagnostic or therapeutic use)

contact: support@agrisera.com

Agrisera AB | Box 57 | SE-91121 Vännäs | Sweden | +46 (0)935 33 000 | www.agrisera.com

Membrane permeabilization (10 min): 0.5% TritonX-100 in PBS (3x5min PBS wash) Blocking (10mins): 5% Fish gelatin in PBS
Primary antibody incubation (2.5 h at 24 °C): Agrisera (AS16 3690) rabbit anti-Cas9 polyclonal antibody diluted 1:100 in blocking buffer (4x5 min blocking buffer wash)
Secondary antibody incubation (1h at 24 °C): goat anti-rabbit DyLight 550 antibody (Agrisera [AS11 1782](#)) diluted 1:300 in blocking buffer. (3x5min PBS wash)
Nuclear counterstaining (5 min): 200ng/ml DAPI in PBS (brief PBS wash)
Mounting: Fluoromount G mounting medium was used to mount coverslips onto glass slides. Imaging: Olympus FV1000 confocal microscope with



40x (NA1.3) oil immersion objective.
BiP localization in 4 days old *Arabidopsis thaliana* roots. BiP signal shown in red, Co-staining with DAPI visualized nucleus (blue color). The material has been fixed in para-formaldehyde for 30 minutes. Tissue cleaning has been performed before immunolocalization. Rabbit anti-BiP primary antibody diluted in 1: 400 and goat anti-rabbit IgG, pre-adsorbed secondary antibody DyLight@550 conjugated (Agrisera) have been used. Scale bar – 10 μm.

Courtesy Dr. Taras Pasternak, Freiburg University